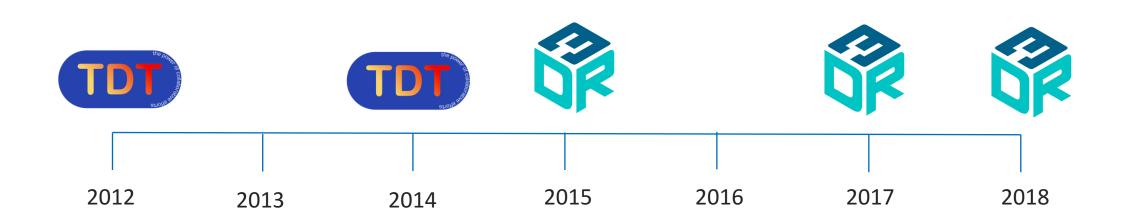


RELAY THERAPEUTICS

How can we get better at this?

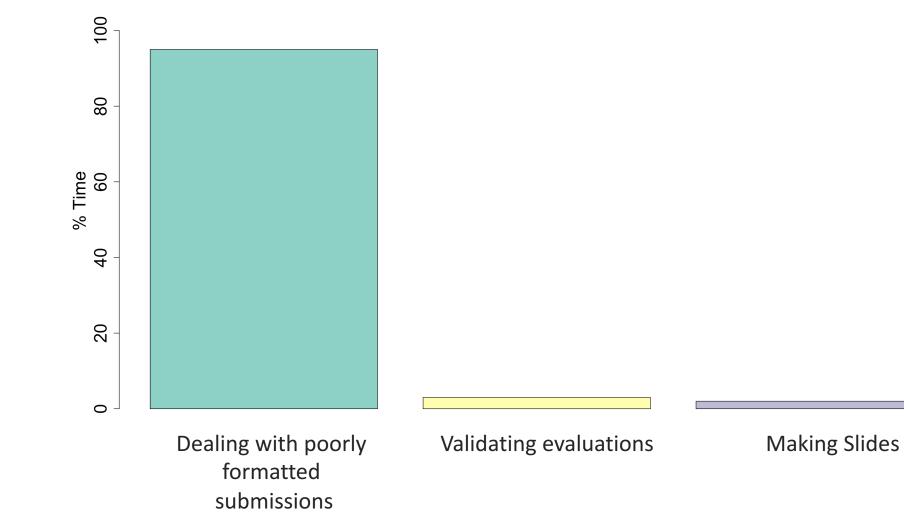
Pat Walters – D3R Workshop February 23, 2018



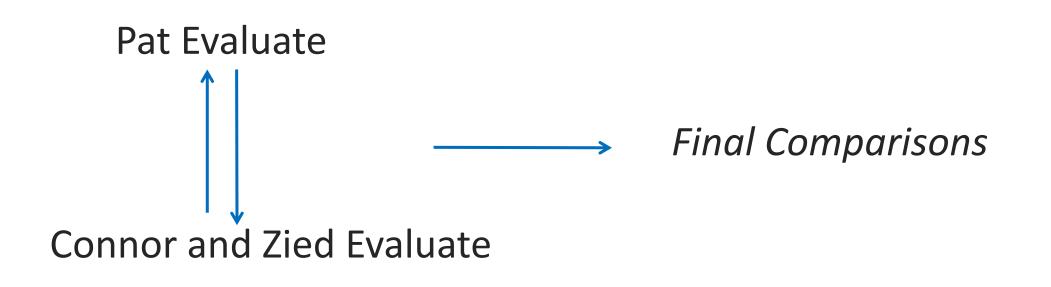


How I Spend My Time On Challenges



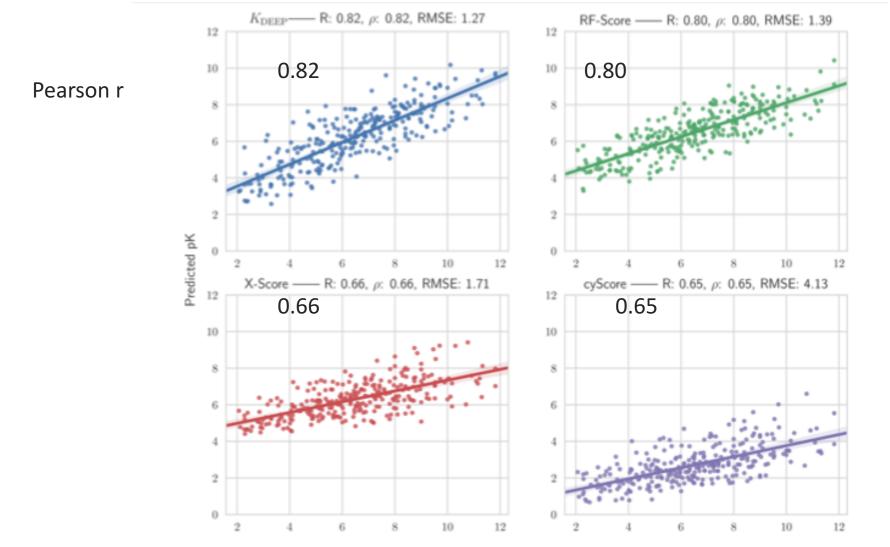






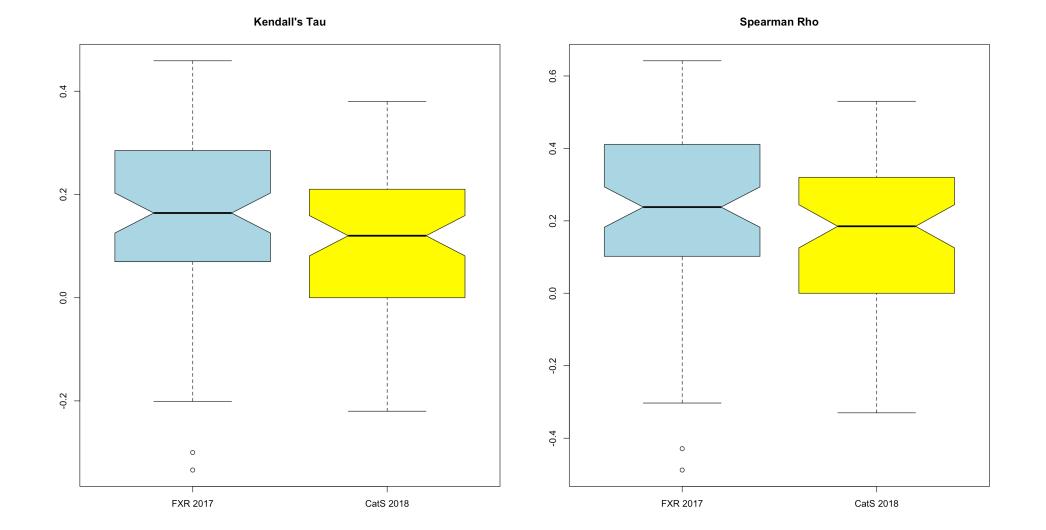
The Literature Makes It Look Like Activity Prediction is a Solved Problem





Experimental pK





6



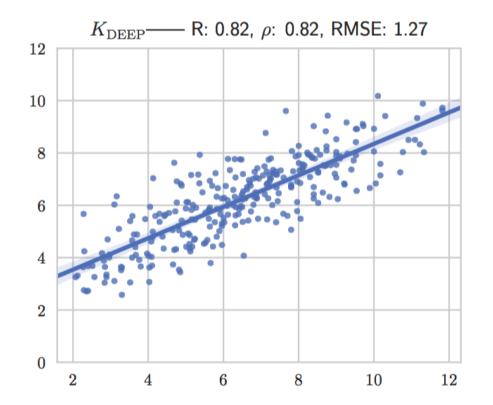
- What constitutes a reasonable dataset
- How data should be reported
- Evaluation metrics
- Statistics for comparison
- What constitutes a null model
- Format of supporting material
- Criteria for reproducibility



- What constitutes a reasonable dataset
- How data should be reported
- Evaluation metrics
- Statistics for comparison
- What constitutes a null model
- Format of supporting material
- Criteria for reproducibility



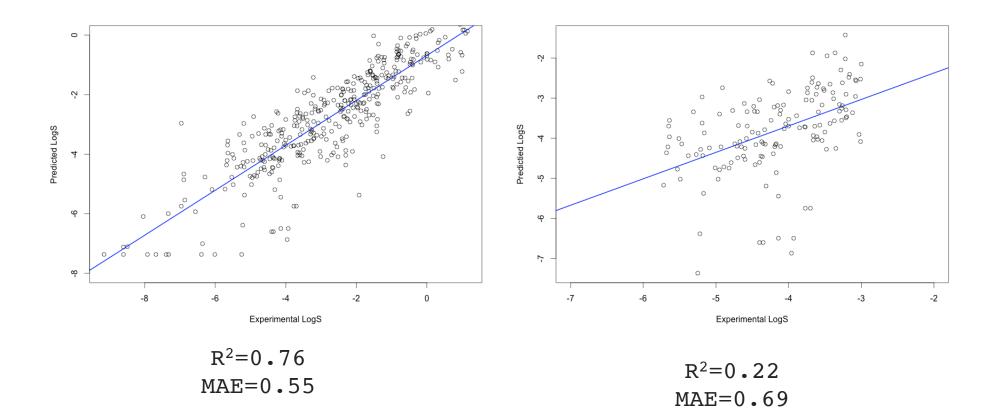
When evaluating a regression model, the dataset should have a dynamic range similar to those observed in drug discovery projects (typically 4-6 logs)



This dataset (PDBind v.2016 core set) spans 10 logs and doesn't provide an appropriate representation of correlation

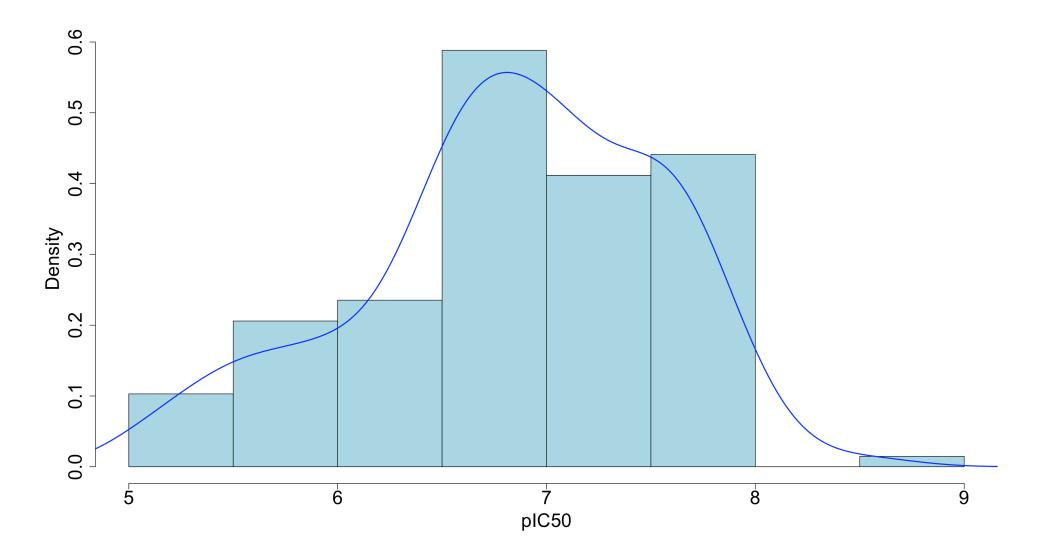


This is the same dataset. On the left we consider the entire set, which has an unrealistically large (~10 log) dynamic range. On the right we consider a more realistic subset with a 3 log dynamic range. Note the change in correlation.



GC3 CatS Dataset Spans a Realistic Dynamic Range

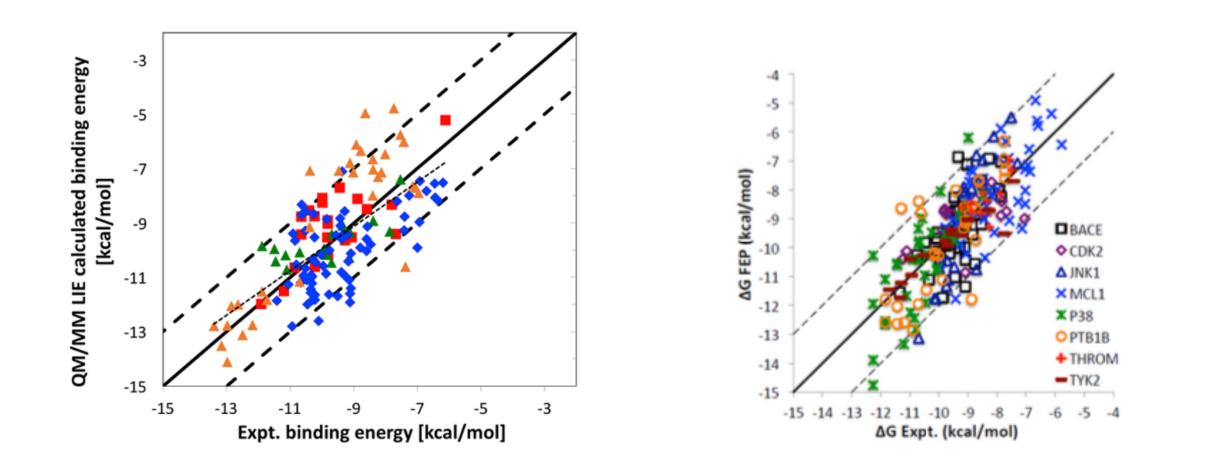






- What constitutes a reasonable dataset
- How data should be reported
- Evaluation metrics
- Statistics for comparison
- What constitutes a null model
- Format of supporting material
- Criteria for reproducibility

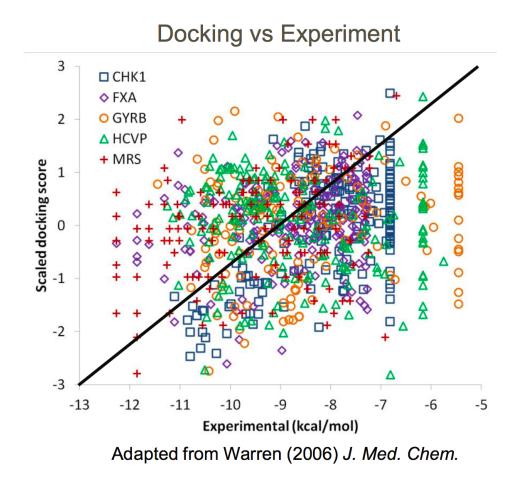




http://pubs.acs.org/doi/abs/10.1021/acs.jpcb.7b07224

http://pubs.acs.org/doi/abs/10.1021/ja512751q

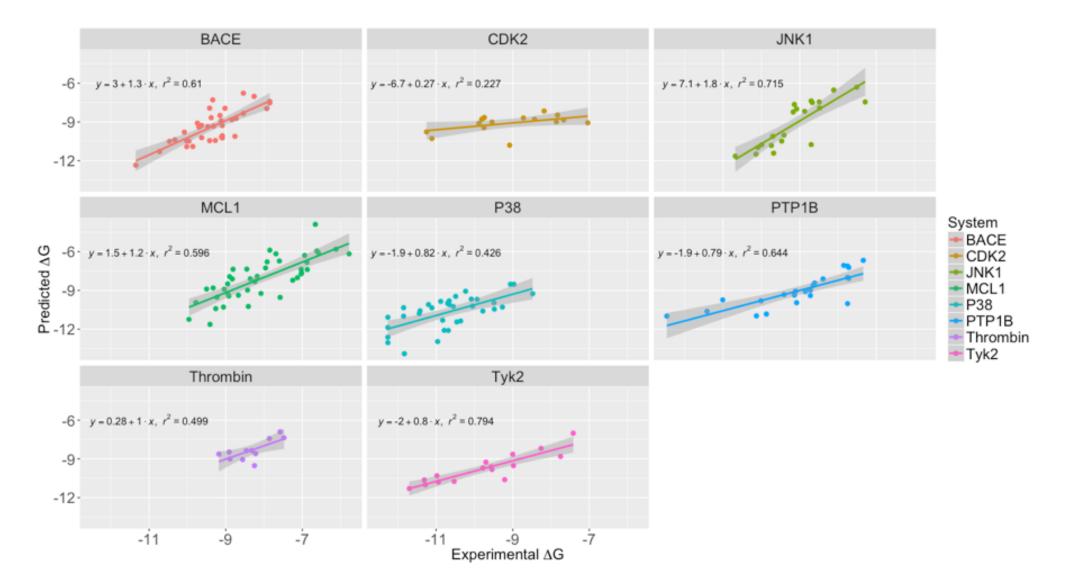




Mill and Neysa (Yesterday)

Trellising provides a much more effective means of comparing datasets







- What constitutes a reasonable dataset
- How data should be reported
- Evaluation metrics
- Statistics for comparison
- What constitutes a null model
- Format of supporting material
- Criteria for reproducibility



Report Pearson, Spearman and Kendall correlations Favor R² over R when reporting a Pearson correlation coefficient Report MAE and/or RMSE

Figure 3. QM/MM LIE calculated binding energy (kcal/mol) vs experimental binding energy (kcal/mol) for BACE1 (red squares), HSP90 (blue diamonds), PERK (orange triangles), and TYK2 (green triangles). The best-fitted line (dotted line), which has a correlation between measured and calculated values of 0.69, has slope = 0.82 and intercept = -1.79.

I have no idea what this means

http://pubs.acs.org/doi/abs/10.1021/acs.jpcb.7b07224



Start with experimental data

Add Gaussian error

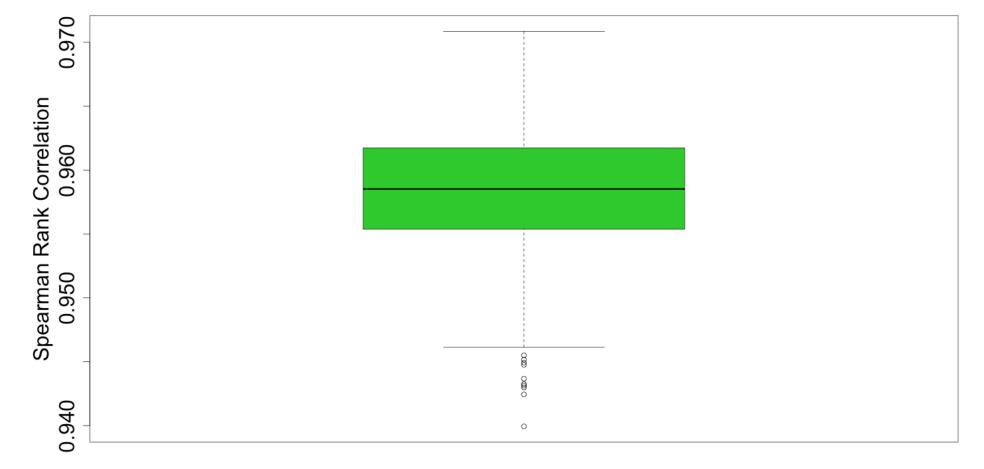
- Mean = 0.0
- Standard deviation = 0.3 log

Calculation correlation

Repeat 1000 times

Brown, Scott P., Steven W. Muchmore, and Philip J. Hajduk. "Healthy skepticism: assessing realistic model performance." *Drug Discovery Today* 14.7 (2009): 420-427.

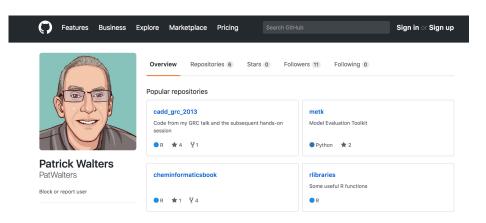




Maximum Achievable Correlation



https://github.com/PatWalters/metk



metk

Model Evaluation Toolkit

In metk, I've collected a set of routines for evaluating predictive models. I put a lot of this code together when I was doing the evaluation for the TDT and D3R projects, as well as a book chapter I wrote in 2013. I'm releasing this project as a way for the community to collaborate and (hopefully) agree on best practices for model evaluation. Most of the initial release is oriented toward the evaluation of free energy calculations.

This is just a start and I plan to add a lot more. Currently, there are routines to calculate

- · Root mean squared (RMS) error
- Mean absolute error (MAE)
- · Pearson correlation coefficient (with confidence limits)
- · Spearman rank correlation (rho) (still need to add confidence limits)
- · Kendall tau (still need to add confience limits)
- Maximum possible correlation given a specific experimental error. This is based on on a 2009 paper by Brown, Muchmore and Hajduk

Most of the statistics is done with routines from scikitlearn and scipy.

The toolkit also includes code to generate a few diagnositc plots that I find helpful when looking at model performance. Examples of these plots can be found here

- A scatter plot of experimental vs predicted ΔG. Lines are drawn at 1 and 2 kcal error
- A histogram of the error distribution.
- The two plots above with ΔG converted to a binding affinity (in uM or nM). On the scatter plot, lines are drawn at 5-fold and 10-fold error. I find that I mentally relate to a fold error in binding affinity better than I do to error expressed in kcal/mol. However, if you like looking at error in kcal/mol, use that plot.

Ultimately, the plan is to implement a number of other methods for model evaluation including those described in papers by Anthony Nicholls.



- What constitutes a reasonable dataset
- How data should be reported
- Evaluation metrics
- Statistics for comparison
- What constitutes a null model
- Format of supporting material
- Criteria for reproducibility

Ensure That Differences in Correlation Are Significant



The molecular mechanics Poisson-Boltzmann surface area (MM-PB/SA) method has bee popular for computing protein-ligand binding free energies in recent years. All previous evaluations of the MM-PB/SA method are based upon computer-generated conformationa ensembles which may be affected by the defective computational methods used fo preparing these conformational ensembles. In an attempt to reach more convincing we have evaluated the MM-PB/SA method on a set of 24 diverse protein-ligan conclusions. complexes, each of which has a set of conformations derived from NMR spectroscopy. Our results indicate that both MM-PB/SA and molecular mechanics generalized Born surface area (MM-GB/SA) are able to produce a modest correlation between their results and the experimentally measured binding free energies on our test set. In particular, both MM-PB/SA and MM-GB/SA produced better results by using a representative structure (rather than averaging over the conformational ensemble of each given comp 0.61-0.74). A head-to-head comparison with four selected scoring functions (X-Score ChemScore, and DrugScore) on the same test set reveals that MM-PB/SA and MM-GB/S results are marginally better than those produced by scoring funcitons, supporting the of the MM-PB/SA method. Nevertheless, scoring functions are still more cost-effective options, especially for high-throughput tasks.

1682

J. Chem. Inf. Model. 2010, 50, 1682-1692

Test MM-PB/SA on True Conformational Ensembles of Protein-Ligand Complexes

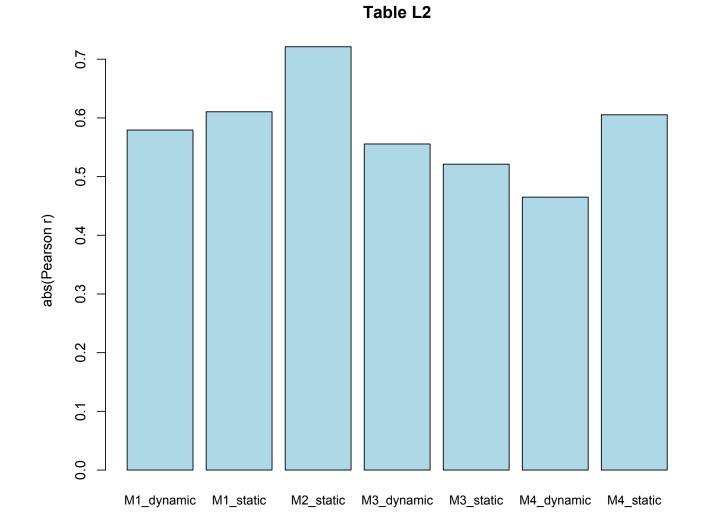
Yan Li, Zhihai Liu, and Renxiao Wang*

State Key Laboratory of Bioorganic Chemistry, Shanghai Institute of Organic Chemistry, Chinese Academy of Science, 345 Lingling Road, Shanghai 200032, People's Republic of China

Received January 25, 2010

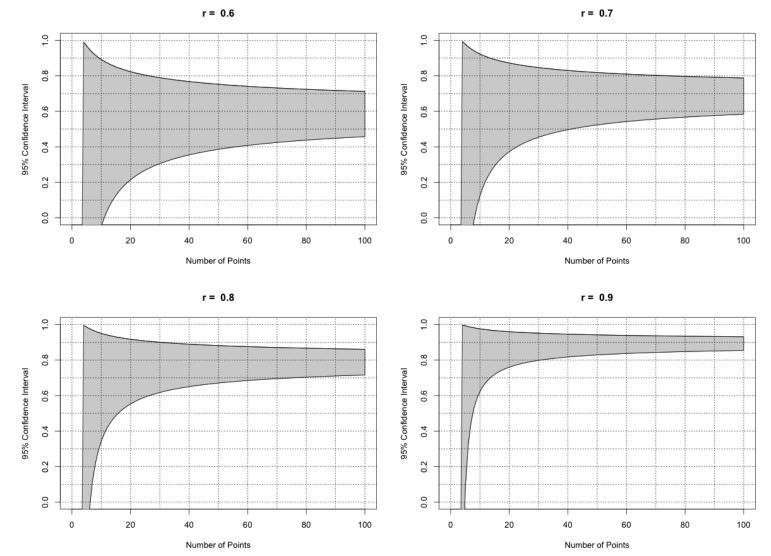
In particular, both MM-PB/SA and MM-GB/SA produced better results by using a representative structure (R) 0.72-0.79) rather than averaging over the conformational ensemble of each given complex (R) 0.61-0.74





A literature comparison of 7 methods for scoring protein-ligand interactions

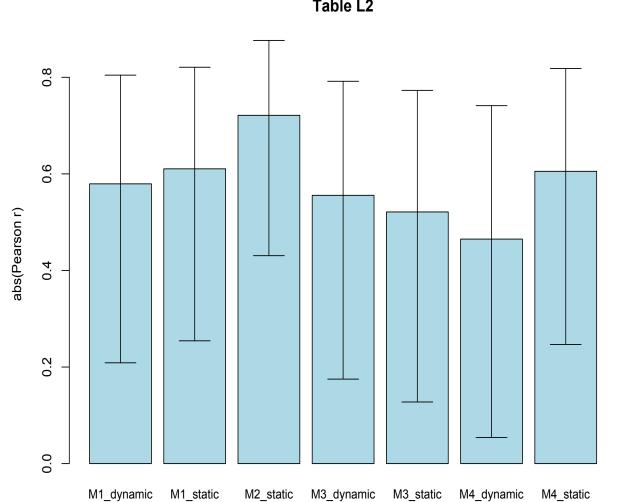
Remember that correlations have confidence intervals and report these intervals



Confidential | © 2017 Relay Therapeutos

It's All the Same!



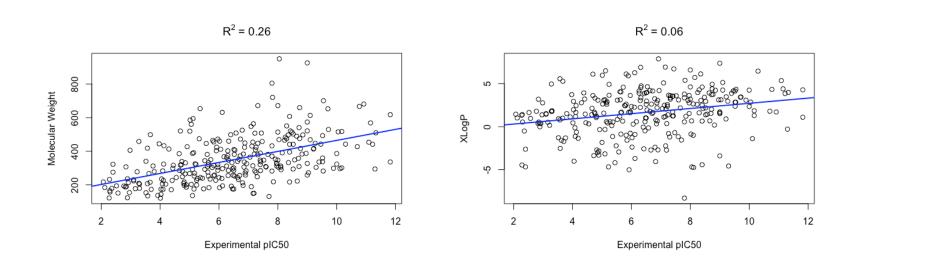






- What constitutes a reasonable dataset
- How data should be reported
- Evaluation metrics
- Statistics for comparison
- What constitutes a null model
- Format of supporting material
- Criteria for reproducibility





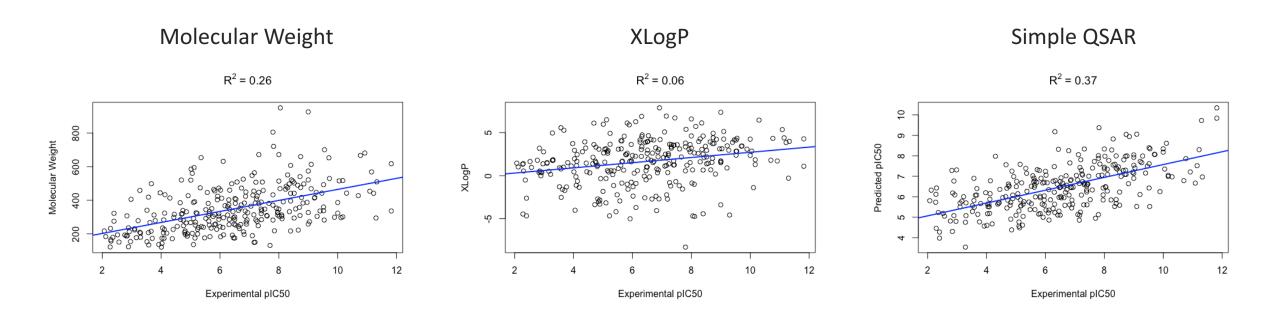


Generate RDKit fingerprints for ligands Train on PDB bind refined set (n=4057) Test on PDB bind core set (n=290) Wall clock time < 5 min

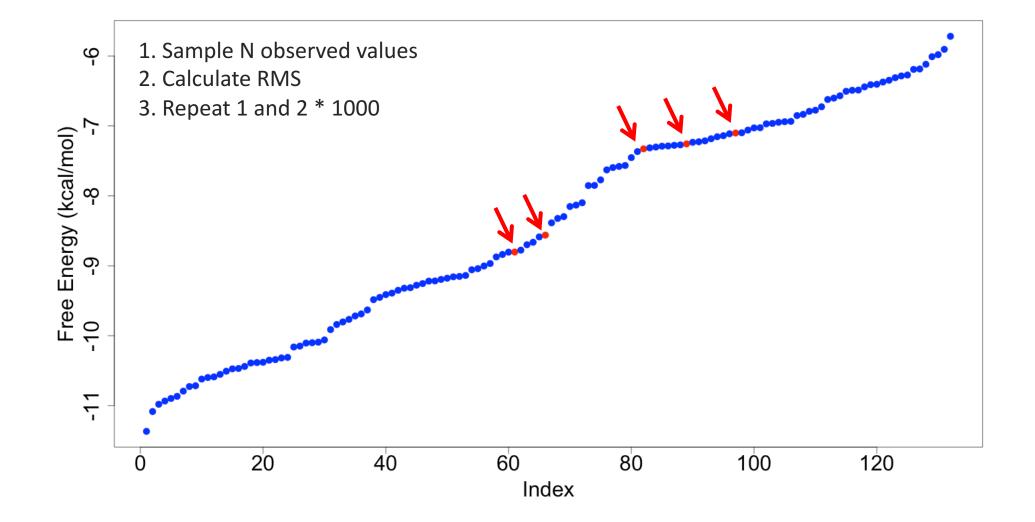
> Predicted pIC50 ω ~ $^{\circ}$ O \circ Experimental pIC50

 $R^2 = 0.37$



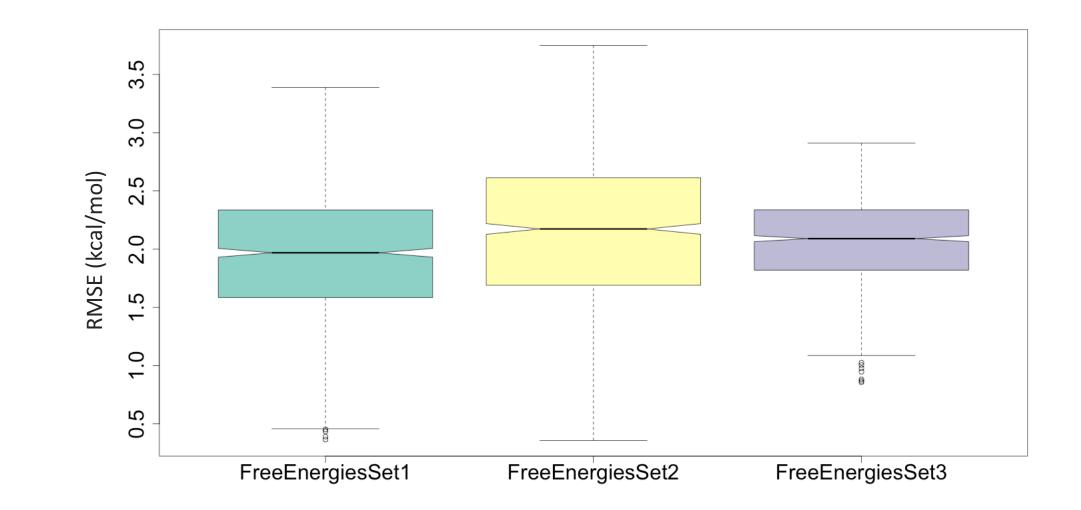






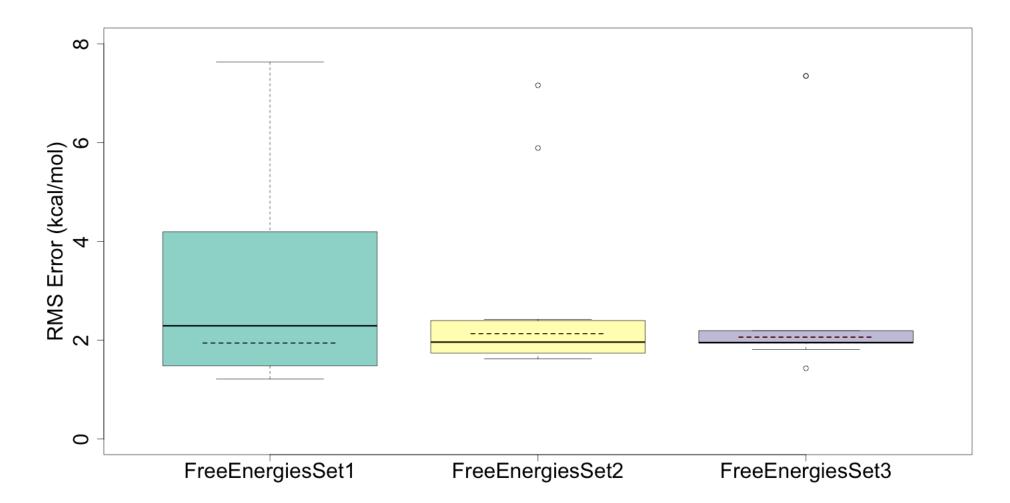
Null Model for GC1 HSP90 Free Energy Challenge







Dashed line indicates the null model





- What constitutes a reasonable dataset
- How data should be reported
- Evaluation metrics
- Statistics for comparison
- What constitutes a null model
- Format of supporting material
- Criteria for reproducibility



Always provide a machine readable table (e.g. csv) of predicted and experimental values

A table in a paper is not sufficient, it is often very difficult to extract tables from pdf files

Chemical structures should be included as SDF or, where appropriate, SMILES to facilitate comparison with other methods

Need to enable readers to evaluate correlations and errors



- What constitutes a reasonable dataset
- How data should be reported
- Evaluation metrics
- Statistics for comparison
- What constitutes a null model
- Format of supporting material
- Criteria for reproducibility



Letter

Modeling, Informatics, and the Quest for Reproducibility

W. Patrick Walters*

Vertex Pharmaceuticals, Inc., 130 Waverly St., Cambridge, Massachusetts 02139, United States

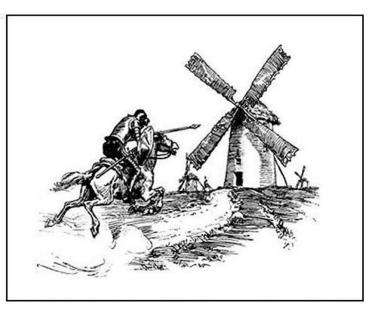
J. Chem. Inf. Model., **2013**, *53* (7), pp 1529–1530 **DOI:** 10.1021/ci400197w Publication Date (Web): June 12, 2013 **Copyright © 2013 American Chemical Society**

*E-mail: pat_walters@vrtx.com Phone: (617) 341-6242.

ACS AuthorChoice - Terms of Use









Code !!!

- A thorough description of your method
- A web implementation

None of the above



We need to agree on

- What constitutes a reasonable dataset
- How data should be reported
- Evaluation metrics
- Statistics for comparison
- What constitutes a null model
- Format of supporting material
- Criteria for reproducibility

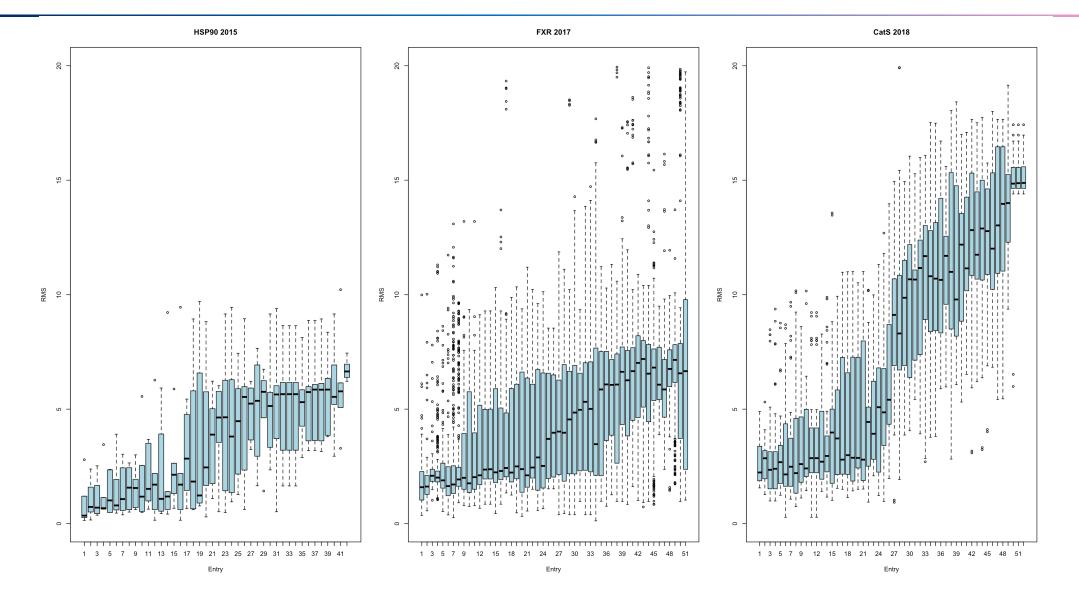
How Can You Help?





Docking Challenges Have Become More Challenging







Are we spending enough time understand compounds that docked poorly?

- Insufficient conformational sampling
- Insufficient pose sampling
- Inadequate scoring
- Ligand poses with limited density

Is everyone missing the same compounds?

Can groups work together to improve their methods?



D3R Participants CSAR Participants TDT Participants SAMPL Participants

Rommie Amaro

Mike Gilson

Mill Lambert

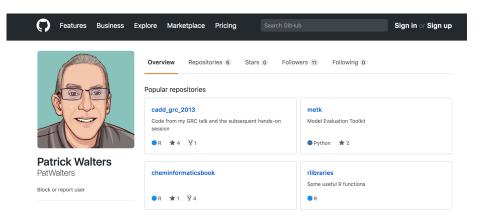
Neysa Nevins

Connor Parks Zied Gaieb

Shuai Liu



https://github.com/PatWalters/metk



metk

Model Evaluation Toolkit

In metk, I've collected a set of routines for evaluating predictive models. I put a lot of this code together when I was doing the evaluation for the TDT and D3R projects, as well as a book chapter I wrote in 2013. I'm releasing this project as a way for the community to collaborate and (hopefully) agree on best practices for model evaluation. Most of the initial release is oriented toward the evaluation of free energy calculations.

This is just a start and I plan to add a lot more. Currently, there are routines to calculate

- · Root mean squared (RMS) error
- Mean absolute error (MAE)
- · Pearson correlation coefficient (with confidence limits)
- Spearman rank correlation (rho) (still need to add confidence limits)
- Kendall tau (still need to add confience limits)
- Maximum possible correlation given a specific experimental error. This is based on on a 2009 paper by Brown, Muchmore and Hajduk

Most of the statistics is done with routines from scikitlearn and scipy.

The toolkit also includes code to generate a few diagnositc plots that I find helpful when looking at model performance. Examples of these plots can be found here

- A scatter plot of experimental vs predicted ΔG. Lines are drawn at 1 and 2 kcal error
- A histogram of the error distribution.
- The two plots above with ΔG converted to a binding affinity (in uM or nM). On the scatter plot, lines are drawn at 5-fold and 10-fold error. I find that I mentally relate to a fold error in binding affinity better than I do to error expressed in kcal/mol. However, if you like looking at error in kcal/mol, use that plot.

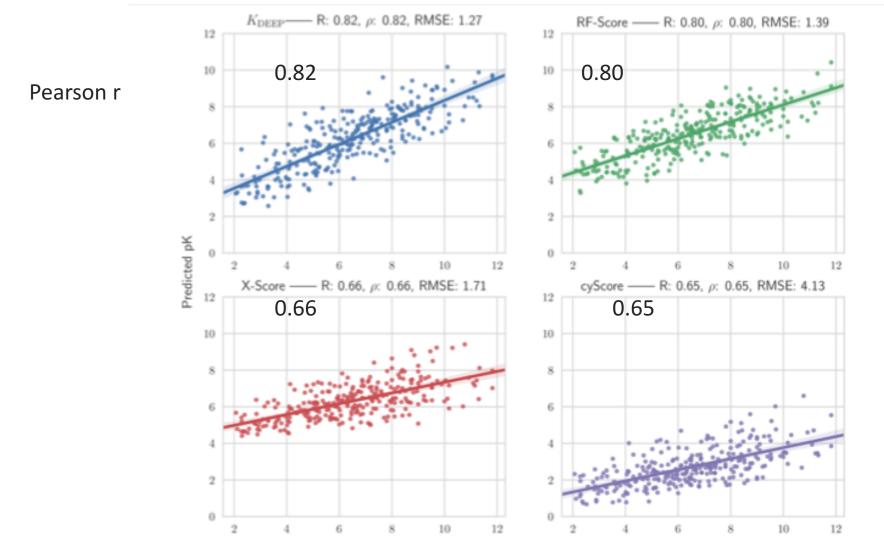
Ultimately, the plan is to implement a number of other methods for model evaluation including those described in papers by Anthony Nicholls.



BACKUP

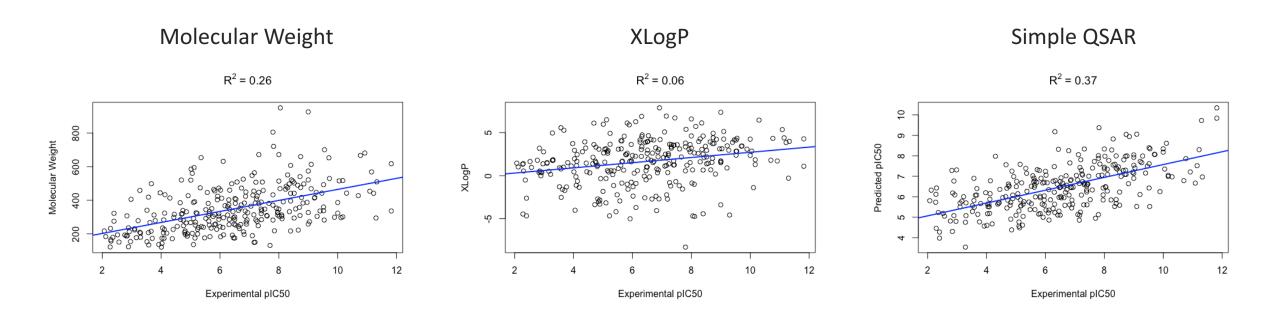
Looks Like Activity Prediction is a Solved Problem



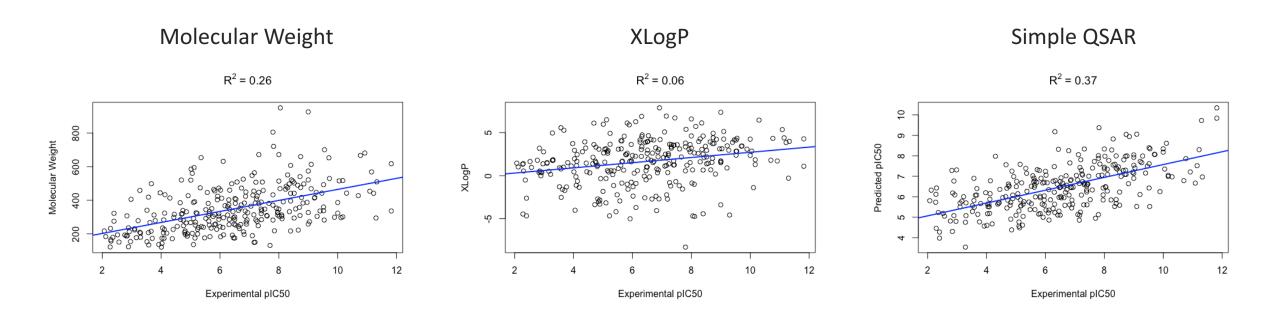


Experimental pK











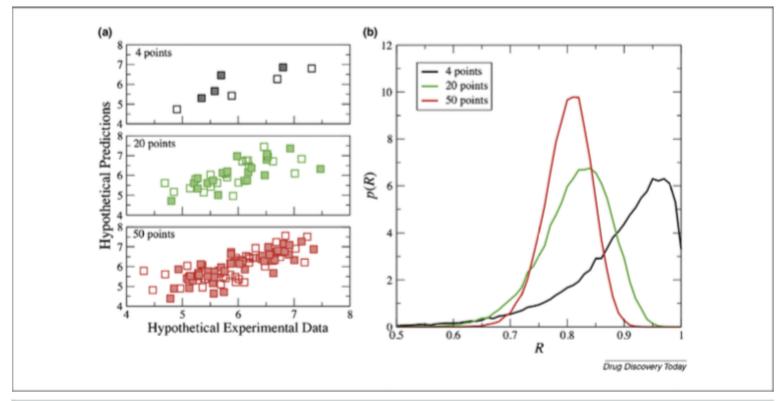


FIGURE 2

Plots illustrating the procedure for introducing sampling error into simulated data. (a) Two independently sampled snapshots (open and filled squares) are shown for three different numbers of points initially distributed over 2 log units. (b) Distribution of possible *R*-values generated by the 'snapshots' of the data in (a). 50,000 iterations were used to generate the *R*-value distributions.

https://www.sciencedirect.com/science/article/pii/S1359644609000403



Start with experimental data

Add Gaussian error

- Mean = 0.0
- Standard deviation = 0.3 log

Calculation correlation

Repeat 1000 times