





and Molecular Biology

D3R Grand Challenge 2

Free Energy Perturbation calculations to predict relative binding affinities for FXR ligands

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27 March 2017

Free Energy Perturbation (FEP) calculations

Zwanzig's formula in complex and in solvent: $\Delta G(A \to B) = G_B - G_A = -kT \ln \left\langle \exp \left(-\frac{V_B - V_A}{kT}\right) \right\rangle_A$ (MBAR used in practice)



 ΔG_1 and ΔG_2 are the free energies of **transfer** of A and B from the unbound to the bound state 2 ΔG_A and ΔG_B are the free energy differences of the **mutation of A into B** in solvent and bound to protein

Molecular dynamics - REST

Molecular Dynamics

Time evolution of the system (Desmond/FEP+, Schrödinger)

Allows for fully flexible receptor — Advantage over docking



Perturbation is achieved with a λ schedule

REST (Replica Exchange with Solute Tempering)

Usual problem in FEP calculations:

- Inability of convergence
- Need for long computational time

Solution:

 Increase of effective temperature to overcome energetic barriers

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• Replica exchange between neighboring λ windows



"thermodynamic axis" alchemical transformation

Issues for consideration before FEP

- The input structure has to be of sufficiently high quality
- Sufficiently long simulations and methodology to overcome barriers (sampling)
- Cannot change the ligand charge during a mutation
- Sensitive to force field (scoring)
- Examination of buried waters (WaterMap, Schrödinger)
- Ensure perturbations are not too big (normally up to 10 heavy atoms)
- Error of the method ~1 kcal/mol
- Large-scale protein movements cannot be sampled sufficiently within the timeframe of FEP calculations

Setting up FEP calculations

This methodology was followed for both spiros and sulfonamides subsets

1) All ligands must belong in the same congeneric series

2) Choice of reference ligand✓ It has to be representative of the series

3) Alignment of all ligands into the reference ligand
✓ After this step, a minimization of the complexes is usually needed

• The input structure has to be of sufficiently high quality: Examples follow with predicted structure and with real crystal structure

<u>Spiros group example</u>

Docking based alignment



Maximum common substructure alignment



Setting up FEP calculations

4) If double occupancy is plausible, both binding modes should be considered in the calculations

- ✓ If they rapidly inter-convert during the simulation, the same $\Delta\Delta G$ is expected, and we can just ignore one of them.
- ✓ If one pose is significantly less stable, discount it from the results
- ✓ If both compounds maintain separate binding poses, but result to the same binding free energy, we can correct the binding free energy for multiple poses (Joseph *et al.*, JCTC, 2015).



Setting up FEP calculations

5) Cannot change the ligand charge during a mutation

✓ We chose to calculate both the sulfonamides and spiros subset in their neutral forms in order not to change the charge.

6) FEP Mapper (FEP+)

- Initial ligands structures as described above are imported
- Ligands are connected through edges based on chemical & binding mode similarity, preservation of ring structure
- \checkmark User can define the cycles
- Every molecule must be part of at least one closed thermodynamic cycle
- Each edge represents the bound & unbound perturbations



Raw errors are predicted from bootstrapping & analytical error of the BAR free energy Cycle closure errors are calculated though the hysteresis of each cycle 7

Running FEP calculations

- ✓ Building of final system geometry
- ✓ OPLS3 force field assignment

✓ Equilibration

- Brownian dynamics with restraints on solute heavy atoms (NVT, T = 10 K, 100 ps, force constant = 50 kcal/mol/Å²)
- MD simulation with restraints on solute heavy atoms (NVT, T = 10K, 12 ps)
- MD simulation with restraints on solute heavy atoms (NPT, 36 ps)
- ✓ MD simulation with no restraints (240 ps)

✓ Production Run

✓ REST MD simulation (NPT, 5 ns)

✓ FEP analysis



System Size

Spiros: 21,000 atoms Sulfonamides: 17,000 atoms

Ligand Conformation Analysis



The force field torsional energy profile (blue curves) are shown superimposed with the probability from the FEP calculations in the complex (solid) and solvent (hashes) simulations for each of the two rotatable bonds in the ligands.

Example of simulation interaction diagram (SID) from FEP calculation



Waters bridges

Comparison of interactions with receptor residues between the two ligands.

Ligand Interaction Diagram (LID) for each of the ligands with residues contacting the ligands and the percent of the simulation spent making each interaction.

$$=$$
 XR_98IC_{50} = 13.1 uM $=$ XR_49IC_{50} = 100 uM10

Exchange density of FEP replicas over λ windows

Solvent Leg

Complex Leg



For both legs of the FXR_98 to FXR_49 simulation, each replica is color coded and the plot shows how it occupies different λ windows during the course of the simulation.

Free energy convergence

Solvent Leq



The total free energy differences between the two ligands (Δ G in kcal/mol) in solvent and complex legs are plotted as a function of time. Three plots for each leg show the accumulated data during different time window schemes; forward; reverse; and sliding window. The tables report the associated bootstrap and analytical errors estimates from corresponding simulation legs.

Spiros series results

#LigandID	Predicted 1st stage	Predicted 2nd stage	Experimental	IC50 (uM)
FXR_10	0	0	0	5.64
FXR_12	-1.51	-3.58	-2.73	0.058
FXR_38	1.53	0.5	1.71	100
FXR_41	-1.84	-2.08	1.71	100
FXR_73	1.24	0.69	0.41	11.2
FXR_74	-1.93	-3.69	-1.28	0.655
FXR_75	1.75	2.94	1.71	100
FXR_76	2.41	-0.41	1.19	41.8
FXR_77	0.18	-4.34	-1.86	0.25
FXR_78	1.74	-3.66	-3.16	0.0283
FXR_79	3.62	0.31	-0.18	4.15
FXR_81	1.21	-4.03	-0.44	2.69
FXR_82	0.35	-3.05	-2.05	0.18
FXR_83	-2.59	-4.61	-1.69	0.33
FXR_84	-0.07	-1.65	-0.13	4.54
FXR_85	-0.02	-2.79	-1.75	0.297
FXR_88	-2.23	-4.39	-1.40	0.54
FXR_89	1.15	-0.14	-1.21	0.735

1st stage



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- Slope improves with crystal structure
- 85% true positives
- Only 2/13 false positives

Sulfonamides series results

#LigandID	Predicted 1st stage	Predicted 2nd stage	Experimental	IC 50 (uM)
FXR_17	0	0	0.00	0.785
FXR_45	-1.05	0.01	2.15	28.9
FXR_46	-0.5	3.41	2.61	62.4
FXR_47	2.99	4.12	1.96	21
FXR_48	0.72	1.19	2.89	100
FXR_49	-0.06	3.34	2.89	100
FXR_91	4.35	5.59	2.16	29.6
FXR_93	5.55	5.2	2.44	46.7
FXR_95	-0.4	3.17	2.21	32.2
FXR_96	-0.63	1.23	2.57	58.9
FXR_98	-2.17	2.83	1.68	13.1
FXR_99	2.35	4.36	2.89	100
FXR_100	-0.05	3.37	1.90	19.1
FXR_101	5.5	2.16	2.12	27.6
FXR_102	5.21	2.21	2.16	29.2





• Narrow range of experimental binding free energies

• **100% success rate** in predicting less active binders than the reference compound (true negatives) 14

FEP conclusions

- FEP+ is predictive given a good initial structure
- A specific protocol has to be followed:
 - \checkmark ligand alignment to a reference ligand structure
 - $\boldsymbol{\checkmark}$ investigation of buried waters
 - $\boldsymbol{\checkmark}$ carefully selecting the mutations
 - $\boldsymbol{\checkmark}$ investigation of double occupancy
- Cannot change the charge during a FEP mutation
- Correlations should not be expected for a narrow range (1-3 kcal/mol) of experimental binding free energies because the error of FEP is ~1 kcal/mol
- High success rate in classifying true positives and true negatives
- Reasonable throughput for lead optimization
 - ✓ 18 spiros, 21,000 atoms, 28 edges, 22 GPUs (Tesla K40m), 29 h
 - ✓ 15 sulfonamides, 17,000 atoms, 19 edges, 22 GPUs, 20 h







D3R Grand Challenge 2

Physics-based pose predictions guided by native ligands

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36 structures for pose prediction



28 relevant PDB crystal structures available

The crystal structure were clustered based on the co-crystallized inhibitor structure:

- Isoxazole derivatives
- Steroid derivative
- Benzimidazole derivatives
- Indole derivatives
- Others
- APO crystal structure was provided by the D3R group
- Wide binding pocket

• Not all ligands can be docked to the same crystal structure

3RUT 3P89 3RUU 3P88 3RVF 3HC6 3HC5 3GD2 3DCT 3FXV 3DCU 4QE6 3BEJ 1OSV 1OT7 4QE8 4OIV 1OSH 3OLF 3OMK 3OMM 3OOF 3OOK 3OKH 3OKI 3L1B 3FLI



Methodology

1) Pose prediction for compounds with known chemotype in crystal structures

- Choice of crystal structure according to known chemotype
- Water molecules that were persistent in crystal structures were kept
- Ligand docking (Glide)
- Alignment to the native ligand (Maestro)
- Minimization of the complex (Maestro)

In case of double occupancy possibility:

- Water thermodynamics in binding pocket (WaterMap)
- Binding pose metadynamics (Desmond)
- FEP calculations (FEP+)





Methodology

2) Pose prediction for compounds with unknown chemotype in crystal structures

- Choice of crystal structure based on
 - a) Shape similarity with native ligands (SHAPE)
 - b) Cross docking in all 28 crystal structures (xglide.py)
 - c) Interaction fingerprints (Maestro)
- · Water molecules that were persistent in crystal structures were kept
- Docking (Glide)
- Alignment with the native ligand when a common core was present (Maestro)
- Minimization of the structure in case of alignment (Maestro)

In case of double occupancy possibility:

Metadynamics calculations were used

Isoxazoles: FXR_4, FXR_23, FXR_33 Sulfonamides: FXR_15-17 Spiros: FXR_10-12 Miscellaneous: FXR_1-3, FXR_18



Pose prediction for compounds with known chemotypes

Benzimidazoles were categorized based on choice of crystal structure

• 30KI

1) Saturated ring

FXR_6, FXR_7, FXR_8, FXR_9, FXR_13, FXR_19, FXR_20, FXR_22, FXR_26, FXR_30, FXR_31, FXR_32, FXR_35

• 30LF

1) Benzene ring, 2) Ortho substituted FXR_14, FXR_24, FXR_25, FXR_27, FXR_28

• 300F

1) Benzene ring, 2) Non ortho substituted FXR_21, FXR_29, FXR_36



3OK



FXR_9

1) Docking in 3OKI

Not consistent binding mode

2) Alignment to 3OKI ligand

3) Minimization of the complex: aligned ligand - protein



Alleviation of clashes

Orange: native ligand Green: docked ligand Orange: native ligand Green: aligned ligand to native Orange: native ligand Green: aligned ligand, complex minimization



FXR_9 (1ytut)

ASN297 SER359 SER336 ILE356 ARG335 TYR373

Orange: crystal structure Green: predicted pose

RMSD = 0.316 Å





FXR_9 (1ytut)

RMSD = 0.316 Å



Receipt ID

Pose prediction for compounds with unknown chemotypes

Isoxazoles

Alignment of isoxazoles



The isoxazole ring is located at the active site quite differently in the released crystal structures (*orange*: FXR_4, *yellow*: FXR_23, *pink*: FXR_33).

FXR 4





FXR_4 (orange) aligned with 30MK native ligand (green) and Prime minimized. Fingerprints Similarity 0.46



Binding pose metadynamics of FXR_4 (orange) aligned with 300F native ligand (green) and Prime minimized. Fingerprints Similarity 0.58



title: FXR_4_protein 300F_aligned____title: 300F_chainA_ligand







FXR_4 (1pdbc)

RMSD = 6.96 Å



Receipt ID

Sulfonamides

FXR 17

XGlide was used and FXR_17 docking pose in 3FLI had the best overlap and the best interaction fingerprints with the native ligand. The docking pose was optimized in Jaguar and re-docked (SP and XP).



Interactions similarity: 0.614







RMSD = 1.63 Å



Receipt ID

Pale color indicates an incomplete set of predictions Green bar indicates your predictions (requires login)

Spiros

Spiros

Shape Results

• Spiros compounds were initially docked in **30MM**, which was indicated by SHAPE analysis.

 Subsequently, they were docked in all 28 crystal structures (cross docking).
The pose of docking in **3FXV** crystal structure was the best, with a Glide Score of ~ -11 kcal/mol.

	10	11	12
30MM	0.538	0.458	0.526
30KH	0.486	0.46	0.43
3P89	0.366	0.366	0.388
3RVF	0.39	0.39	0.38
10SH	0.490	0.524	0.450
40IV	0.550	0.466	0.507
4QE8	0.550	0.466	0.507
3L1B	0.40	0.40	0.40
1077	0.56	0.64	0.56





Orange: crystal structure Green: predicted pose ASN297 HIS451 ARG335 **SER336 TYR373** Docking in 3OMM – 2nd pose Docking in 3FXV – 1st pose

RMSD = 2.14 Å

RMSD = 4.43 Å



Receipt ID

Green bar indicates your predictions (requires login)

Conclusions

- Pose predictions were accurate, when a crystal structure with common chemotype native ligand was available. In this case, docking, alignment to native ligand and minimization performed well.
- Methodology needs improvements, in case a crystal structure with common chemotype native ligand is not available.
 In this case, cross docking and interaction fingerprints performed well for some compounds.
- Difficulty in predicting isoxazoles poses due to diversity in binding modes

Acknowledgements

Dr. Stephan Ehrlich Dr. Thomas Steinbrecher

D3R organizers

Cournia lab members, Biomedical Research Foundation, Academy of Athens

http://www.drugdesign.gr/

SCHRÖDINGER.



